

Molecular Diagnostics

REAGENTS FOR
OLIGONUCLEOTIDE
LABELING

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1. Introduction

Toolbox for oligonucleotide labeling

ELITechGroup® has been actively involved in the development of fluorescence-based nucleic acid probe technologies since the early 1990s. This experience has helped us better understand the challenges of nucleic acid probe manufacturing and the needs of end users. The proprietary reagents and techniques that we have been developing have two main goals in mind: first, to provide labeled probes with the desired physicochemical properties, and, second, to simplify the probes' synthesis and purification. To achieve these goals, we have put together a comprehensive toolbox for nucleic acid labeling which includes a wide range of unique reagents:

- **Fluorescent dyes** for signal detection
- **Quenchers** for signal control
- **Nucleic Acid Base Analogues (Super Bases)** for modulation of nucleic acid duplex stability
- **Universal Nucleic Acid Bases** to control hybridization specificity
- **Minor Groove Binders (MGBs)** to enhance DNA duplex stability and hybridization specificity
- **Intercalators** for modulation of nucleic acid duplex stability
- **Artificial Nucleic Acids** for orthogonal nucleic acid technologies
- **Lipophilic groups** for therapeutic nucleic acid applications
- **Spacers and Linkers** to assemble various nucleic acid probe components together and optimize their performance.

These are offered in a variety of reagent forms:

- **2-Cyanoethyl Phosphoramidites** for internal and 5'-end labeling
- **Synthesis solid supports** for 3'-end labeling
- **Activated esters** for post-synthesis conjugation with aminecontaining substrates

In addition, all of our Super Bases are available as both 3'- and 5' phosphoramidites to make them compatible with both standard $(3' \rightarrow 5')$ and 'reverse' (5' \rightarrow 3') DNA synthesis approaches. The reverse approach offers additional synthesis flexibility, for instance, in situations when a synthesis solid support is only available for 5'-end labeling.

Toolbox applications: DNA probe-based detection chemistries

Numerous DNA detection technologies that rely on fluorophore-labeled nucleic acids have been developed in the last several decades. Simple endlabeled fluorescent probes are useful in heterogeneous assays wherein the target-probe duplex is separated from the excess probe before signal detection (e.g., nucleic acid microarray technologies). On the other hand, most of the nucleic acid amplification technologies (e.g., real-time PCR) are performed in a homogeneous format with no separation of components and require a fluorescence change during or after amplification for target detection. The table below contains examples of some DNA detection chemistries, with the emphasis on ELITechGroup's proprietary MGB technology. More detailed analysis of such chemistries can be found in References [1](#page-210-1) and [2.](#page-210-2) Signal generation for all of these chemistries can generally be attributed to either hydrolysis or hybridization events. The hydrolysis type probes, when annealed to a target, are enzymatically degraded to separate the fluorophore ('Fluor') from a quenching ('Q') moiety, whereas hybridization probes/primers generate fluorescence signal via a hybridization-triggered conformational change. Hybridization probes and primers have the benefit of post-amplification melting curve analysis to confirm the specificity of amplification or detect target mutations.

**TaqMan is a trademark of Roche Molecular Systems, Inc.*

Structurally, the degree of complexity varies among the chemistries. In most cases 5'- or 3'- fluorophore and quencher reagents are involved, as well as 5'- or 3'-MGB for MGB probes. In more complicated cases, internal dyes and spacers are required.

ELITechGroup offers a wide range of dye and MGB reagents for postsynthesis and on-line incorporation to meet the challenges of any of those chemistries. Most of our reagents are available as internal phosphoramidites for internal and multiple dye incorporation. We also offer amine and dye-modified nucleoside phosphoramidites that allow for internal modifications with minimal impact on duplex stability.

MGB TaqMan (Section [9,](#page-145-0) [Figure 13A](#page-148-0)), **DSQ TaqMan** (Sectio[n 12\)](#page-167-0), **TaqMan** probes, **Molecular Beacons,** or **FRET** probes can be synthesized using our quencher synthesis supports (Sections [3,](#page-81-0) [11,](#page-163-0) and [12\)](#page-167-0), standard 3' nucleoside phosphoramidites, and concluded with fluorophore phosphoramidites. Alternatively, **Endonuclease IV** probes, which require a 5'-quencher and a 3'-fluorophore with a special linker to increase mismatch discriminating ability (Ref. [3\)](#page-210-3), are prepared starting from a fluorophore synthesis support followed by 3'-nucleoside phosphoramidites and a quencher phosphoramidite at the last step.

For probes with internal fluorophore or quencher (**Scorpions, LUX primers** or **MGB FRET** (Section [9,](#page-145-0) [Figure 13D](#page-148-0)) we offer fluorophore/quencher internal phosphoramidites with a non-nucleoside linker or nucleoside phosphoramidites with fluorophore/quencher attached to a nucleobase.

To further increase flexibility, reduce the consumption of valuable MGB and fluorophore reagents, and improve product quality in some cases we recommend using 5'-nucleoside phosphoramidites (a.k.a. 'reverse' phosphoramidites) for probe synthesis. For example, **MGB Pleiades** probes (Section [9,](#page-145-0) [Figure 13C](#page-148-0)) contain 5'-MGB, 5'-fluorophore, and 3'-quencher moieties. For this configuration, the most economical synthetic path is to start from a 5'-Fluorophore-MGB synthesis support (Section [13\)](#page-172-0), follow with 5'-nucleoside phosphoramidites for a probe sequence and complete with a quencher phosphoramidite. Having the hydrophobic quencher incorporated at the last step of the synthesis provides an efficient handle for reverse phase HPLC purification. To accommodate this approach with the duplex enhancing **Super Bases** (Section [5\)](#page-116-0) we offer 'reverse' 5' phosphoramidites of the Super Bases as well. Similarly, **MGB Eclipse** probes (Section [9,](#page-145-0) [Figure 13B](#page-148-0)) can be synthesized starting from an EDQ-MGB synthesis support (Section [11\)](#page-163-0), followed by 5'-nucleoside phosphoramidites addition, and a fluorophore phosphoramidite at the last step. M**GB Fluorogenic primers** (Sectio[n 9,](#page-145-0) [Figure 12\)](#page-147-0), which are analogous to MGB Pleiades with an unmodified 3'-hydroxyl end, can also be synthesized using the 'reverse' phosphoramidite approach.

2. Fluorescent Dyes

ELITechGroup's dyes are available in several fluorescent colors ranging from blue to far red. Their excitation and fluorescence emission spectra (Figure 1 and [Figure 2\)](#page-6-1) have been optimized to match filter sets for most commercial fluorescence-based nucleic acid diagnostic instruments. With assay multiplexing in mind, the dyes have also been designed to maximize spectral separation and minimize channel cross-talk.

Figure 1 Absorption spectra of representative fluorophores.

Figure 2 Emission spectra of representative fluorophores.

For most of our fluorophores, we offer three types of labeling reagents: 2 cyanoethyl dye phosphoramidites, dye-modified oligonucleotide synthesis supports for on-line dye incorporation and amine-reactive active esters for post-synthesis conjugation. These three types of reagents are detailed below.

2-Cyanoethyl Phosphoramidites

2-Cyanoethyl dye phosphoramidite chemistry is undeniably one of the most convenient methods for the preparation of labeled oligonucleotides. We offer dye phosphoramidites whenever they are synthetically accessible and sufficiently stable during oligonucleotide synthesis and deprotection. Our internal phosphoramidites contain a multifunctional hydroxyprolinol linker with one primary hydroxyl, which is blocked by the DMT group, one secondary hydroxyl for phosphoramidite incorporation, and an amine for dye attachment via an amide bond. This design allows dye incorporation at any desired position within a probe's sequence. In addition, if used in the last coupling cycle, the DMT group offers a convenient DMT-on purification handle, especially for dyes that do not significantly alter C18 HPLC retention.

Synthetically more accessible terminal phosphoramidites with a straight chain C6 linker are only suitable for single dye incorporation at the end of oligonucleotide synthesis but may be a viable alternative to the internal phosphoramidites depending on probe design and purification requirements.

Oligonucleotide Synthesis Supports

Dye-labeled controlled pore glass (CPG) and polystyrene (PS) supports are very economical methods for dye labeling. Depending on whether 5' or 3' nucleoside phosphoramidites are used, this approach allows essentially quantitative 3' or 5' dye incorporation. All of our dye CPG or PS supports use the proprietary trifunctional hydroxyprolinol moiety, which provides a DMT-blocked primary hydroxyl (the starting point for oligonucleotide synthesis) and a secondary hydroxyl for connecting the dye to the synthesis support via a cleavable succinate linker.

Active esters

Most of our dyes are available as amine-reactive pentafluorophenyl (PFP) esters or fused lactones with activities similar to well-known NHS esters. Both non-aqueous and aqueous/organic conjugation conditions can be used.

Active esters: Post-synthesis conjugation using non-aqueous DMSO conditions

For conjugation applications, one particularly useful property of oligonucleotides is the solubility of their trialkylammonium forms in organic solvents such as DMSO. These forms can be prepared in a variety of ways. For example, a triethylammonium (TEA) form can be obtained by simple C18 reverse phase chromatography (which is usually done prior to a postsynthesis conjugation) using triethylammonium bicarbonate buffer. The volatile buffer can be readily removed by evaporation *in vacuo*. The resultant oligonucleotide (TEA form) is now soluble in DMSO and can be modified with a variety of agents under mostly anhydrous conditions. This approach allows use of conjugation reagents that are either insoluble or unstable in the presence of significant amounts of water, making the conjugation reaction more robust.

Another useful attribute of the non-aqueous protocol is that all oligonucleotide-related material can be easily precipitated by adding a solution of a sodium salt (such as iodide, perchlorate, etc.) in acetone, and separated from the bulk of unreacted dye, thus simplifying conjugate purification.

Overall, our experience indicates that when compared to aqueous/organic conditions, the non-aqueous conjugation procedure is more reproducible; it affords higher coupling yields and simplifies conjugate purification.

Active esters: Two-step conjugation procedures

Preparation of dye-labeled nucleic acid probes by post-synthesis conjugation may present significant challenges in terms of conjugation efficiency and separation of the resultant conjugate from unconjugated dye. The presence of unlabeled oligonucleotide will reduce the probe's quality and ultimately its performance. On the other hand, even minor contamination with unconjugated dye may lead to an undesired increase in background signal.

Conjugates can often be separated from the starting oligonucleotide and unconjugated dye by reverse phase HPLC. It is therefore desirable, for purification purposes, to use fairly hydrophobic dyes that provide significant changes in HPLC retention time. Excessive hydrophobicity, however, may lead to undesired dye aggregation and reduction in the probe's performance. To address this dilemma, we developed a two-step conjugation procedure. In the first step, the main conjugation reaction is done using a transiently protected, hydrophobic form of a dye followed by simple reverse phase HPLC purification. In the second step, the conjugate is treated with a deblocking agent releasing the dye in a more hydrophilic form and purified again. Importantly, the dual HPLC purification procedure simplifies the removal of free, unincorporated dye. This approach is illustrated in [Figure](#page-9-0) 3 for a conjugation reaction between the AquaPhluor 525 lactone (M830683) and an amine-modified oligonucleotide. The AquaPhluor 525 dye, in its fully deprotected form, is highly hydrophilic and provides no useful change in the retention time for HPLC purification. On the other hand, its intermediate has the phosphonate group blocked by the trifluoroacetamidobutyl (TFAAB) groups, which provide a very convenient purification handle. The blocking groups are removed at the next step by treatment with ammonium hydroxide generating the highly polar zwitterionic phosphonate.

Figure 3 Two-step post-synthesis incorporation of AquaPhluor 525 using its active lactone form (AP525 lactone, M830683).

On-line incorporation of AquaPhluor $^{\circledR}$ Dyes

Our proprietary AquaPhluor dyes have been developed for direct on-line incorporation of polar fluorescent dyes into oligonucleotide probes. In their form as labeling reagents, they possess latent phosphonate groups which are fully protected and neutral and, therefore, compatible with the phosphoramidite preparation and coupling chemistries. Upon completion of standard oligonucleotide synthesis, in their ultimate form the dyes are negatively charged and thus significantly more hydrophilic. The phosphonate chemistry of AquaPhluor dyes makes it possible to combine the versatility of automated oligonucleotide synthesis with the benefits of hydrophilicity. As an additional benefit, the phosphonate is a convenient attachment point for a variety of linking and functional groups. Based on how the phosphonate group is utilized, the AquaPhluor dyes can be divided into two classes. AquaPhluor 525, for example, has a phosphonate that is blocked with two TFAAB groups, one of which is eventually removed producing a highly hydrophilic zwitterion [\(Figure](#page-10-0) 4).

Figure 4 Two types of AquaPhluor chemistries for on-line dye incorporation.

In the second class, which includes AP559, AP593, AP570, AP639, AP642, AP662, AP680 and AP690 dyes, a reactive linking group is integrated as one of the phosphonate blocking groups. After incorporation of the dye into an oligonucleotide and treatment with deblocking agents, only the special blocking group is removed leaving the conjugation linkage intact. Both types of the dyes are compatible with phosphoramidite, active ester, and oligonucleotide chemistries.

Epoch Blue 380

Acronym: EB380

Epoch Blue 380 is a UV light-excitable blue fluorescent aminocoumarin dye with excitation and emission properties similar to those of Marina Blue and Alexa Fluor 350. It is fully compatible with oligonucleotide synthesis and deprotection.

 $00 M^{-1}$ cm⁻¹ 200 M⁻¹cm⁻¹ (Epoch Blue 380-HEG) 000 $M⁻¹cm⁻¹$ 0 (moderately hydrophobic) ependent between pH of 5 and 8 (Appendix C)

**Measured for T8-Epoch Blue 380 conjugate (50 mM Tris-HCl pH 8.5, 20°C)*

***Measured for T8-Epoch Blue 380 conjugate (0.1 M Sodium tetraborate)*

****Percent acetonitrile required for elution of T8-EB380-HEG Dye Calibrator (M300585) from C18 column (see Appendix A for details)*

*****Suitable for ABI 3900 DNA synthesizer*

Epoch Blue 380 PFP ester

Product number: M830726; CAS #: 2378004-31-8

**Purity of this active derivative is verified by HPLC & NMR and usually exceeds 90%. Some loss of activity, however, may occur during packaging and storage, especially for small (1- 10 mg) unit sizes.*

Epoch Blue 380 Internal Phosphoramidite

Product number: M830192; CAS #: 2378004-42-1

Epoch Blue 380-HEG Phosphoramidite

Product number: M830727; CAS #: 2378004-26-1

Recommended storage conditions/Stability \leq -10 \degree C, dry/ TBD
Available documents Certificate of anal

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Certificate of analysis

Epoch Blue 380 CPG

Product number: M830669

Epoch Blue 380 Polystyrene

Product numbers: M830670 (bulk), M100441 (columns)

**ABI 3900 type columns, ~200 nmol/column*

AquaPhluor® 492 NHS ester

Product number: M830194; CAS #:2378004-15-8

AquaPhluor 492 is a highly hydrophilic fluorescent dye with excitation and emission properties similar to Alexa Fluor 488 and fluorescein. Compared to fluorescein, AP492 is less pH-sensitive and more photostable.

AP492 NHS ester is suitable for conjugation reactions with aminecontaining substrates.

492 nm 513 nm $13,500 \, \text{M}^{-1} \text{cm}^{-1}$ $73,000 \, \text{M}^{\text{-1}} \text{cm}^{\text{-1}}$ TBD (hydrophilic)

**Measured for T8-AP492 conjugate (50 mM Tris-HCl pH 8.5, 20°C)*

***Measured for T8-AP492 conjugate (0.1 M Sodium tetraborate)*

****Percent acetonitrile required for elution of T8-AP492 from C18 column (Appendix A) ****Purity of this active derivative is verified by HPLC & NMR and usually exceeds 70%. Some loss of activity, however, may occur during packaging and storage, especially for small (1-10 mg) unit sizes.*

6-FAM

FAM (6-Carboxyfluorescein) is a popular green fluorescent dye compatible with standard oligonucleotide synthesis and deprotection.

Absorbance maximum* 496 nm Emission maximum* 517 nm Extinction coefficient (260 nm) $33,300 \text{ M}^1 \text{cm}^{-1}$ (FAM-HEG) Extinction coefficient (496 nm) 80,400 M 1 cm 1 Fluorescence quantum yield**(Φ) 0.88 Hydrophobicity by C18 chromatography*** pH dependence of dye fluorescence pK_a 6.8-7.0 (see Appendix C)

19.2 (hydrophilic)

**Measured for T8-FAM-HEG conjugate (50 mM Tris-HCl pH 8.5, 20°C)*

***Measured for T8-FAM-HEG conjugate (0.1 M Sodium tetraborate)*

****Percent acetonitrile required for elution of T8-FAM-HEG Dye Calibrator (M300586)*

from C18 column (see Appendix A for details)

*****Suitable for ABI 3900 DNA synthesizer*

6-FAM Phosphoramidite

Product number: M100011; CAS #: 204697-37-0

FAM-HEG Phosphoramidite

Product number: M830100; CAS #: 2378004-27-2

FAM-HEG CPG

Product number: M830719

FAM-HEG Polystyrene

Product numbers: M830718 (bulk), M100256 (columns)

**ABI 3900 type columns, ~200 nmol/column*

FAM-dU 3'-Phosphoramidite

Product numbers: M830763; CAS #: 2378004-02-3

FAM-dU 5'-Phosphoramidite

Product numbers: M830764; CAS #: 2378004-07-8

FAM-Super I 3'-Phosphoramidite

Product numbers: M830783; CAS #: 2378004-06-7

Gig Harbor Green™

Acronym: GG

Gig Harbor Green is an alternative to FAM with similar excitation and emission properties.

Gig Harbor Green reagents are compatible with standard DNA synthesis and deprotection conditions.

**Measured for T8-GG conjugate (50 mM Tris-HCl pH 8.5, 20°C)*

***Measured for T8-GG conjugate (0.1 M Sodium tetraborate)*

****Percent acetonitrile required for elution of T8-GG Dye calibrator from C18 column (see Appendix A for details)*

*****Suitable for ABI 3900 DNA synthesizer*

Gig Harbor Green CPG

Product number: M830771

Gig Harbor Green Polystyrene

Product numbers: M400090 (bulk), M100433 (columns)

**ABI 3900 type columns, ~200 nmol/column*

AquaPhluor 525

Acronym: AP525

AquaPhluor 525 is a hydrophilic fluorescent dye with excitation and emission properties similar to VIC, JOE, HEX and Alexa Fluor 532. It possesses a zwitterionic 4-aminobutyl- phosphonate group for increased hydrophilicity and reduced aggregation. The latent phosphonate is conveniently protected in the reactive forms of the dye.

**Measured for T8-AP525 conjugate (50 mM Tris-HCl pH 8.5, 20°C)*

***Measured for T8-AP525 conjugate (0.1 M Sodium tetraborate)*

****Percent acetonitrile required for elution of T8-AP525 Dye Calibrator (M300587) from*

C18 column (see Appendix A for details)

*****Suitable for ABI 3900 DNA synthesizer*

AquaPhluor 525 Lactone

Product number: M830683; CAS #: 2378004-23-8

**Purity of this active derivative is verified by HPLC (via test reaction with an aliphatic amine) & NMR and usually exceeds 90%. Some loss of activity, however, may occur during packaging and storage, especially for small (1-10 mg) unit sizes.*

AquaPhluor 525 Terminal Phosphoramidite

Product number: M830714; CAS #: 876746-10-0

AquaPhluor 525 Internal Phosphoramidite

Product number: M830199; CAS #: 2378004-28-3

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AquaPhluor 525-HEG Phosphoramidite

Product number: M100104; CAS #: 2378004-19-2

AquaPhluor 525 Polystyrene

Product numbers: M830672 (bulk), M100276 (columns)

**ABI 3900 type columns, ~200 nmol/column*

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Yakima Yellow®

Acronym: YY

Yakima Yellow is a fluorescent dye with excitation and emission properties similar to VIC, JOE, HEX and Alexa Fluor 532. It is fully compatible with standard oligonucleotide synthesis and deprotection.

525 nm 547 nm $28.600 \, \text{M}$ ⁻¹cm⁻¹ $86,500 \, M^{-1}$ cm $^{-1}$ 25.6 (moderately hydrophobic) Independent above pH 7 (see Appendix C)

**Measured for T8-YY conjugate (50 mM Tris-HCl pH 8.5, 20°C)*

***Measured for T8-YY conjugate (0.1 M Sodium tetraborate)*

****Percent acetonitrile required for elution of T8-YY Dye Calibrator (M830730) from C18 column (see Appendix A for details)*

Yakima Yellow Lactone

Product number: M830684; CAS #: 502484-58-4

**Purity of this active derivative is verified by HPLC (via test reaction with an aliphatic amine) & NMR and usually exceeds 90%. Some loss of activity, however, may occur during packaging and storage, especially for small (1-10 mg) unit sizes.*

Yakima Yellow Phosphoramidite

Product number: M830552; CAS #: 502485-39-4

Yakima Yellow CPG Product number: M830395

Yakima Yellow Polystyrene

Product numbers: M400063 (bulk), M100443 (columns)

**ABI 3900 type columns, ~200 nmol/column*

AquaPhluor 559

Acronym: AP559

AquaPhluor 559 is a fluorescent dye with excitation and emission properties similar to TAMRA and NED. The dye is compatible with oligonucleotide synthesis and deprotection.

559 nm $28,300 \, M^{-1}$ cm $^{-1}$ $86,500 \, M^{-1}$ cm $^{-1}$ 25.6 (moderately hydrophobic) Weakly dependent between pH of 5 and 8 (see Appendix C)

**Measured for T8-AP559 conjugate (50 mM Tris-HCl pH 8.5, 20°C)*

***Measured for T8-AP559 conjugate (0.1 M Sodium tetraborate)*

****Percent acetonitrile required for elution of T8-AP559 Dye Calibrator (M301430) from C18 column (see Appendix A for details)*

AquaPhluor 559 Terminal Phosphoramidite

Product number: M830698; CAS #: 2378004-43-2

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AquaPhluor 559 Internal Phosphoramidite

Product number: M830195; CAS #: 2378004-38-5

AquaPhluor 559 CPG

Product number: M830673

AquaPhluor 559 Polystyrene

Product numbers: M830674 (bulk), M100278 (columns)

**ABI 3900 type columns, ~200 nmol/column*

AquaPhluor 570

Acronym: AP570

AquaPhluor 570 is a fluorescent dye with excitation and emission properties similar to Redmond Red. Unlike Redmond Red it is pH insensitive and fully compatible with standard oligonucleotide synthesis and deprotection.

Absorbance maximum* F mission maximum $*$ Extinction coefficient (260 nm) Extinction coefficient (570 nm) Fluorescence quantum yield**(Φ) 0.68 Hydrophobicity by C18 chromatography*** pH dependence of dye fluorescence

**Measured for T8-AP570 conjugate (50 mM Tris-HCl pH 8.5, 20°C)*

***Measured for T8-AP570 conjugate (0.1 M Sodium tetraborate)*

****Percent acetonitrile required for elution of T8-AP570 Dye Calibrator (M830700) from*

C18 column (see Appendix A for details)

AquaPhluor 570 Terminal Phosphoramidite

Product number: M830699; CAS #: 2378004-30-7

AquaPhluor 570 Internal Phosphoramidite

Product number: M830675; CAS #: 2378004-20-5

AquaPhluor 570 CPG

Product number: M830676

AquaPhluor 570 Polystyrene

Product numbers: M830677 (bulk), M100444 (columns)

**ABI 3900 type columns, ~200 nmol/column*

Redmond Red®

Acronym: RR

Redmond Red is a fluorescent dye with excitation and emission properties similar to those of AquaPhluor 570. It is compatible with oligonucleotide synthesis.
Mild deprotection conditions are deprotection conditions are recommended.

580 nm 594 nm $9,800 \, M^{-1}$ cm $^{-1}$ $64,500 \, M^{-1}$ cm⁻¹ 17.9 (hydrophilic) Highly pH-dependent, pK_a ~7 (see Appendix C)

**Measured for T8-RR conjugate (50 mM Tris-HCl pH 8.5, 20°C)*

***Measured for T8-RR conjugate (0.1 M Sodium tetraborate)*

****Percent acetonitrile required for elution of T8-RR Dye Calibrator (M830731) from C18 column (see Appendix A for details)*

Redmond Red Lactone

Product number: M830728; CAS #: 502484-16-4

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**Purity of this active derivative is verified by HPLC (via test reaction with an aliphatic amine) & NMR and usually exceeds 90%. Some loss of activity, however, may occur during packaging and storage, especially for small (1-10 mg) unit sizes.*

Redmond Red Phosphoramidite

Product number: M830035; CAS #: 909906-38-3

Redmond Red CPG

Product number: M830390

Redmond Red Polystyrene

Product numbers: M830729 (bulk), M100445 (columns)

**ABI 3900 type columns, ~200 nmol/column*

AquaPhluor 593

Acronym: AP593

AquaPhluor 593 is a fluorescent dye with excitation and emission properties similar to ROX and Texas Red. The dye is
stable under the standard stable under the standard oligonucleotide synthesis and deprotection conditions.

 $\overline{593}$ nm $19.680 \, \text{M}^{-1} \text{cm}^{-1}$ $115,240 \, M^{-1}$ cm $^{-1}$ 29.9 (hydrophobic) Weakly dependent between pH 5 and 8 (Appendix C)

**Measured for T8-AP593 conjugate (50 mM Tris-HCl pH 8.5, 20°C)*

***Measured for T8-AP593 conjugate (0.1 M Sodium tetraborate)*

****Percent acetonitrile required for elution of T8-AP593 Dye Calibrator (M300392) from*

C18 column (see Appendix A for details)

AquaPhluor 593 PFP ester

Product number: M830196; CAS #: 1140966-99-9

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**Purity of this active derivative is verified by HPLC & NMR and usually exceeds 70%. Some loss of activity, however, may occur during packaging and storage, especially for small (1-10 mg) unit sizes.*

AquaPhluor 593 Terminal Phosphoramidite

Product number: M830736; CAS #: 2378004-17-0

AquaPhluor 593 CPG

Product number: M830208

AquaPhluor 593 Polystyrene

Product numbers: M830732 (bulk), M100257 (columns)

**ABI 3900 type columns, ~200 nmol/column*

AquaPhluor 639

Acronym: AP639

AquaPhluor 639 is a fluorescent dye with excitation and emission properties similar to Cy5, Quasar 670 and AP642. It features sequence-independent absorption and fluorescence spectra and excellent chemical and photochemical stability. AP639 is compatible with both standard and mild oligonucleotide deprotection conditions.

Absorbance maximum^{*} Emission maximum* Extinction coefficient (260 nm) Extinction coefficient (639 nm) Fluorescence quantum yield**(Φ) Hydrophobicity by C18 chromatography*** pH dependence of dye fluorescence

**Measured for T8-AP639 conjugate (50 mM Tris-HCl pH 8.5, 20°C)*

***Measured for T8-AP639 conjugate (0.1 M Sodium tetraborate)*

****Percent acetonitrile required for elution of T8-AP639 Dye Calibrator (M301715) from*

C18 column (see Appendix A for details)

AquaPhluor 639 Terminal Phosphoramidite

Product number: M830741; CAS #: 2378724-63-9

AquaPhluor 639 CPG

Product numbers: M830757

AquaPhluor 639 Polystyrene

Product numbers: M830743 (bulk), M100440 (columns)

**ABI 3900 type columns, ~200 nmol/column*

AquaPhluor 642

Acronym: AP642

AquaPhluor 642 is a fluorescent dye with excitation and emission properties similar to Cy5. Mild deprotection conditions are recommended for products carrying this dye.

Absorbance maximum* E mission maximum $*$ Extinction coefficient (260 nm) Extinction coefficient (641 nm) Fluorescence quantum yield $**$ (Φ) Hydrophobicity by C18 chromatography*** pH dependence of dye fluorescence

**Measured for T8-AP642 conjugate (50 mM Tris-HCl pH 8.5, 20°C)*

***Measured for T8-AP642 conjugate (0.1 M Sodium tetraborate)*

****Percent acetonitrile required for elution of T8-AP642 Dye Calibrator (M300589) from C18 column (see Appendix A for details)*

AquaPhluor 642 PFP ester

Product number: M830197; CAS #: 2378043-93-5

**Purity of this active derivative is verified by HPLC & NMR and usually exceeds 80%. Some loss of activity, however, may occur during packaging and storage, especially for small (1-10 mg) unit sizes.*

AquaPhluor 642 CPG

Product number: M830190

AquaPhluor 642 Polystyrene

Product numbers: M830678 (bulk), M100280 (200 nmol column), M100279 (1 µmol column)

**ABI 3900 type columns*

AquaPhluor 662

Acronym: AP662

AquaPhluor 662 is a fluorescent dye with excitation and emission properties similar to those of Cy5. Mild deprotection conditions are recommended for products carrying this dye.

 $6.600 \, M^{-1}$ cm⁻¹ $1.100 \, \text{M}^{\text{-1}} \text{cm}^{\text{-1}}$.0 (hydrophobic) dependent between pH 5 and 8 (Appendix C)

**Measured for T8-AP662 conjugate (50 mM Tris-HCl pH 8.5, 20°C)*

***Measured for T8-AP662 conjugate (0.1 M Sodium tetraborate)*

****Percent acetonitrile required for elution of T8-AP662 Dye Calibrator (M830724) from*

C18 column (see Appendix A for details)

AquaPhluor 662 PFP ester

Product number: M830198; CAS #: 2378043-92-4

**Purity of this active derivative is verified by HPLC & NMR and usually exceeds 85%. Some loss of activity, however, may occur during packaging and storage, especially for small (1-10 mg) unit sizes.*

AquaPhluor 662 CPG

Product number: M830191

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AquaPhluor 662 Polystyrene

Product numbers: M830679 (bulk), M100271 (columns)

**ABI 3900 type columns, ~200 nmol/column*

AquaPhluor 680

Acronym: AP680

AquaPhluor 680 is a fluorescent dye with excitation and emission properties similar to those of Cy5.5. Mild deprotection conditions are recommended for products carrying this dye.

Absorbance maximum* 682 nm Emission maximum* 704 nm Extinction coefficient (260 nm) $16,100M^{-1}cm^{-1}$ Extinction coefficient (682 nm) 221,300 M⁻¹cm⁻¹ Fluorescence quantum yield**(Φ) 0.07 Hydrophobicity by C18 chromatography*** 35.8** (hydrophobic) pH dependence of dye fluorescence Independent between pH 5 & 8

(Appendix C)

**Measured for T8-AP680 conjugate (50 mM Tris-HCl pH 8.5, 20°C)*

***Measured for T8-AP680 conjugate (0.1 M Sodium tetraborate)*

****Percent acetonitrile required for elution of T8-AP680 Dye Calibrator (M300753) from*

C18 column (see Appendix A for details)

*****Suitable for ABI 3900 DNA synthesizer*

AquaPhluor 680 PFP ester

Product number: M830682; CAS #: 2378004-18-1

**Purity of this active derivative is verified by HPLC & NMR and usually exceeds 90%. Some loss of activity, however, may occur during packaging and storage, especially for small (1-10 mg) unit sizes.*

AquaPhluor 680 CPG

Product number: M830680

AquaPhluor 680 Polystyrene

Product numbers: M830681 (bulk), M100282 (columns)

**ABI 3900 type columns, ~200 nmol/column*

AquaPhluor 690

Acronym: AP690

AquaPhluor 690 is a fluorescent dye with excitation and emission properties similar to AP680, Cy5.5 and Quasar 705. It features sequence-independent absorption and fluorescence spectra and improved chemical and photo-chemical stability as compared to the spectrallysimilar AquaPhluor 680, Cy5.5 and Quasar 705 dyes.

Absorbance maximum* 689 nm Emission maximum* 709 nm Extinction coefficient (260 nm) $23,500 \text{ M}^{\text{-1}}\text{cm}^{\text{-1}}$ Extinction coefficient (689 nm) $141,900 \text{ M}^{-1} \text{cm}^{-1}$ Fluorescence quantum yield**(Φ) 0.14 Hydrophobicity by C18 chromatography*** 30.5 (hydrophobic) pH dependence of dye fluorescence Independent between pH 5 & 8 (Appendix

C)

**Measured for T8-AP690 conjugate (50 mM Tris-HCl pH 8.5, 20°C)*

***Measured for T8-AP690 conjugate (0.1 M Sodium tetraborate)*

****Percent acetonitrile required for elution of T8-AP690 Dye Calibrator (M302123) from*

C18 column (see Appendix A for details)

*****Suitable for ABI 3900 DNA synthesizer*

AquaPhluor 690 Terminal Phosphoramidite

Product number: M830760; CAS #: 2378004-21-6

AquaPhluor 690 CPG

Product numbers: M830759

AquaPhluor 690 Polystyrene

Product numbers: M830755 (bulk), M100500 (columns)

**ABI 3900 type columns, ~200 nmol/column*

3. Fluorescence Quenchers Azo Dye Quenchers

ELITechGroup's azo dye quenchers have absorption spectra covering fluorophores with emissions from 400 nm to 750 nm [\(Figure 5\)](#page-81-0). They have been designed with the relatively long range (20-60 Å) fluorescence resonance transfer (FRET) quenching mechanism in mind, which requires a spectral overlap between fluorophore emission and quencher absorption spectra.

Figure 5 Absorption spectra of azo dye quenchers: Quencher-470 (blue), Eclipse[®] Dark Quencher (pink), Quencher-575 (green), Quencher-630 (red).

Azo dyes are also known to exert non-FRET quenching, so-called contact quenching. Contact quenching (also known as quenching by 'touching' or collisional quenching) is not based on the long range FRET mechanism and requires close contact between fluorophore and quencher. This mechanism works especially well in Molecular Beacons wherein 'touching' is enhanced by the formation of the self-complementary double-stranded stem.

Based on the dynamics of fluorophore-quencher interactions, quenching was categorized as being either dynamic or static in complex formation (Ref. [4\)](#page-210-0). Three different dynamic contact quenching mechanisms were identified: intersystem crossing, electron exchange, and photoinduced electron transfer (PET). At least the last two mechanisms have been shown to be present in known nucleic acid probes. For example, the electron

exchange (also known as Dexter interaction) is present (along with FRET) in linear dual labeled probes (such as TaqMan) and requires temporary orbital overlap. The photoinduced electron transfer between a fluorescent dye and one or more guanine bases, located in close proximity to the dye, is the quenching mechanism for DNA probes described in Ref. [5](#page-210-1) and [6.](#page-210-2) Examples of PET-mediated quenching by nitroindole nucleosides are also described (Ref. [7\)](#page-210-3). It has been shown (Ref. [8\)](#page-210-4) that the CDPI3-type MGB demonstrates the PET mechanism as well. An example of static quenching is described in Ref. [9;](#page-210-5) it is characterized by the formation of a ground state complex (hetero-dimer) between a fluorophore and an azo dye quencher accompanied by significant changes in absorption spectra of the dyes.

Contact quenching is present to a varying degree in other probe chemistries that do not have any intentional secondary structure (e.g., TaqMan). For such probes the contribution of contact quenching will generally diminish as the length of the probe increases.

In general, when selecting a quencher, one should consider which quenching mechanism is predominant for a given probe chemistry. For example, for longer non-MGB TaqMan probes, wherein FRET is expected to be the main quenching mechanism, a quencher with the best spectral overlap with fluorophore emission should be selected. On the other hand, for MGB TaqMan or MGB Pleiades probes, which are generally shorter and also benefit from the MGB-induced PET quenching, the Eclipse Dark Quencher will work well regardless of fluorophore emission spectrum.

Lite Quenchers™

Lite Quenchers (LQ-380 and LQ-400) are nitrodiarylethene (nitrostilbene) analogues (Ref. [10\)](#page-210-6) whose absorption spectra are substantially blue-shifted relative to the emission spectra of common fluorophores (such as fluorescein). These so-called non-FRET quenchers do not rely on the spectral overlap of quencher absorbance and fluorophore emission (FRET mechanism) for their quenching abilities. They have been designed to eliminate (or significantly weaken) the FRET and preserve the contact quenching. As a result, the oligonucleotide probes possessing such quenchers have fluorescence characteristics that are not always achievable using traditional FRET quenchers.

Lite Quenchers are particularly useful in oligonucleotide hybridization probes having fewer than 12 bases between the fluorophore and quencher. Oligonucleotide probes of this length, when labeled with traditional FRET quenchers, demonstrate significantly reduced fluorescence signals upon

hybridization with their targets. The signal drop is due to insufficient spatial separation between the fluorophore and quencher leading to a residual FRET quenching. The Lite Quenchers, which have substantially diminished FRET quenching properties, avoid this disadvantage.

A particular class of probes that is suitable for use in conjunction with the Lite Quenchers are very short (8-12 bases long) MGB Pleiades probes. Such probes are short due to the duplex stabilizing effect of the MGB and are optimal for distinguishing closely related nucleic acid targets. Importantly, these probes containing the Lite Quenchers demonstrate low background fluorescence and high hybridization signals, the latter being significantly higher than those achieved using the traditional FRET-based quenchers. Both background fluorescence and signal are essential parameters for improving assay sensitivity. Such probes are useful, for instance, in digital PCR, such as described in Ref[. 11.](#page-210-7)

Molecular Beacons are another type of probe that may demonstrate improved fluorescence characteristics when used with Lite Quenchers.

Azo dye and Lite Quenchers are available as 2-cyanoethyl dye phosphoramidites, dye-modified oligonucleotide synthesis solid supports for on-line dye incorporation, and amine-reactive esters for post-synthesis conjugation.

Lite Quencher-380

Acronym: LQ-380

LQ-380 is a contact quencher with no spectral overlap with fluorophores emitting above 500 nm.

Products containing LQ-380 quencher are fully compatible with standard oligonucleotide synthesis and deprotection conditions.

**Measured for T8-LQ-380 conjugate (50 mM Tris-HCl pH 8.5, 20°C)*

***Suitable for ABI 3900 DNA synthesizer*

Lite Quencher-380 PFP ester

Product number: M830787; CAS #: 2244400-57-3

**Purity of this active derivative is verified by HPLC & NMR and usually exceeds 90%. Some loss of activity, however, may occur during packaging and storage, especially for small (1-10 mg) unit sizes.*

Lite Quencher-380 Internal Phosphoramidite

Product number: M830786; CAS #: 2244400-74-4

Lite Quencher-380 CPG

Product number: M830799

Lite Quencher-380 Polystyrene

Product numbers: M830793 (bulk), M100505 (columns)

**ABI 3900 type columns, ~200 nmol/column*

Lite Quencher-400

Acronym: LQ-400

LQ-400 is a contact quencher with a minimal spectral overlap with the emission of fluorescein and no-overlap with fluorophores emitting above 525 nm.

Mild deprotection conditions are recommended for products containing LQ-400 quencher.

**Measured for T8-LQ-400 conjugate (50 mM Tris-HCl pH 8.5, 20°C)*

***Suitable for ABI 3900 DNA synthesizer*

Lite Quencher-400 PFP ester

Product number: M830789; CAS #: 2244400-58-4

**Purity of this active derivative is verified by HPLC & NMR and usually exceeds 90%. Some loss of activity, however, may occur during packaging and storage, especially for small (1-10 mg) unit sizes.*

Lite Quencher-400 Internal Phosphoramidite

Product number: M830788; CAS #: 2244400-75-5

Lite Quencher-400 CPG

Product number: M830800

Lite Quencher-400 Polystyrene

Product number: M830794 (bulk), M100507 (columns)

**ABI 3900 type columns, ~200 nmol/column*

Quencher-470

Acronym: Q-470

Quencher-470 is a Dabcyl-based nonfluorescent quencher. Its principal application is a quencher for blue and green fluorescent donor dyes for FRET probes. The quencher is fully compatible with standard oligonucleotide synthesis and deprotection.

**Measured for T8-Q470 conjugate (50 mM Tris-HCl pH 8.5, 20°C)*

***Suitable for ABI 3900 DNA synthesizer*

Quencher-470 PFP ester

Product number: M830572; CAS #: 2378004-22-7

**Purity of this active derivative is verified by HPLC & NMR and usually exceeds 90%. Some loss of activity, however, may occur during packaging and storage, especially for small (1-10 mg) unit sizes.*

Quencher-470 Internal Phosphoramidite

Product number: M830565; CAS #: 2378004-32-9

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Quencher-470 CPG

Product number: M830562

Quencher-470 Polystyrene

Product numbers: M830568 (bulk), M100446 (columns)

**ABI 3900 type columns, ~200 nmol/column*

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Eclipse Dark Quencher

Synonyms: Eclipse Quencher, EDQ

Eclipse dark quencher is a universal nonfluorescent quencher for FRET probes. The quencher is fully compatible with standard oligonucleotide synthesis and deprotection.

**Measured for T8-Eclipse Quencher conjugate (50 mM Tris-HCl pH 8.5, 20°C)*

***Suitable for ABI 3900 DNA synthesizer*

Eclipse Dark Quencher PFP ester

Product number: M830573; CAS #: 2378004-34-1

**Purity of this active derivative is verified by HPLC & NMR and usually exceeds 90%. Some loss of activity, however, may occur during packaging and storage, especially for small (1-10 mg) unit sizes.*

Eclipse Dark Quencher Phosphoramidite

Product number: M830028; CAS #: 2378004-24-9

Eclipse Dark Quencher Polystyrene

Product numbers: M830486 (bulk), M100447 (columns)

**ABI 3900 type columns, ~200 nmol/column*

EDQ-Super I 3'-Phosphoramidite

Product number: M830781; CAS #: 2378004-12-5

Quencher-575

Acronym: Q-575

 $N_{\stackrel{>}{\scriptstyle\sim} N}$ N $NO₂$ Cl $\frac{1}{\alpha}$ $OCH₃$ H_3 CO

Quencher-575 quencher is a universal nonfluorescent quencher for FRET probes. The quencher is compatible with standard oligonucleotide synthesis. Mild deprotection is recommended.

**Measured for T8-Q575 conjugate (50 mM Tris-HCl pH 8.5, 20°C)*

***Suitable for ABI 3900 DNA synthesizer*

Quencher-575 PFP ester

Product number: M830574; CAS #: 2378004-39-6

**Purity of this active derivative is verified by HPLC & NMR and usually exceeds 90%. Some loss of activity, however, may occur during packaging and storage, especially for small (1-10 mg) unit sizes.*

Quencher-575 Internal Phosphoramidite

Product number: M830566; CAS #: 2378004-36-3

Quencher-575 Polystyrene

Product numbers: M830569 (bulk), M100448 (columns)

**ABI 3900 type columns, ~200 nmol/column*

Quencher-630

Acronym: Q-630

Quencher-630 is the most red-shifted quencher currently available from ELITechGroup, and it is especially suitable for quenching red and far red fluorophores including the AP680 dye. The quencher is compatible with standard oligonucleotide synthesis. Mild deprotection is recommended.

**Measured for T8-Q630 conjugate (50 mM Tris-HCl pH 8.5, 20°C)*

***Suitable for ABI 3900 DNA synthesizer*

Quencher-630 PFP ester

Product number: M830575; CAS #: 2378004-33-0

**Purity of this active derivative is verified by HPLC & NMR and usually exceeds 90%. Some loss of activity, however, may occur during packaging and storage, especially for small (1-10 mg) unit sizes.*

Quencher-630 Internal Phosphoramidite

Product number: M830567; CAS #: 2378004-29-4

Quencher-630 CPG

Product number: M830564

Quencher-630 Polystyrene

Product numbers: M830570 (bulk), M100449 (columns)

**ABI 3900 type columns, ~200 nmol/column*

4. Dye Calibrators

Dye calibrators are available for evaluation of our fluorescent dyes' properties and instrument calibrations. The calibrators are T_8 conjugates with the dyes attached to the 3' terminus via the hydroxyprolinol linker. All conjugates are carefully purified and formulated as 0.1 mM solution in 1x TE buffer.

 \pm 0.5% of calculated molecular weight (MW) \leq -10°C, protect from light 3 years at \leq -10°C Certificate of analysis

*All absorption and emission data were obtained in 50 mM Tris-HCl pH 8.5 at 20°C. *Acetonitrile concentration (%) required to elute a dye calibrator from C18 column (see Appendix A for details).*

5. Nucleic Acid Base Analogues

In the field of molecular diagnostics, modified nucleobases are often utilized to modulate hybridization properties of natural nucleic acids (NAs). Short NA fragments are used as probes to identify specific NA targets and as primers to amplify these targets. Depending on the application, it may be necessary to adjust duplex stability and specificity. For example, degeneracy of the genetic code, target polymorphism, and a high rate of pathogen mutagenesis often impart strict requirements on assay design, necessitating very short probes and amplification primers. In this case, the hybridization strength of natural NA bases may not be sufficient to satisfy the assay requirements and, therefore, demand the use of some hybridization-enhancing techniques. For other applications, it may be imperative to ignore sequence variations (polymorphisms), which would require overcoming the fundamental specificity of the natural A:T and G:C base-pairs. ELITechGroup has developed a number of artificial NA base analogues to help fulfill those requirements.

Super G®

Super G (8-aza-7-deazaguanosine) demonstrates approximately the same base pairing specificity as natural G. However, unlike natural G, due to the absence of the N-7 nitrogen atom, it cannot form non-canonical base pairs. As a result, Super G effectively eliminates undesired secondary structures inherent to natural G-rich sequences and improves hybridization efficiency. The Super G analogue is recommended for use in NA probes and primers to disrupt G stretches.

An additional benefit of Super G for fluorogenic hybridization probes is its low fluorescence quenching tendency, which is an intrinsic and often undesired property of natural G. The resultant Super G-modified probes demonstrate higher fluorescence efficiency and, therefore, assay sensitivity.

Super A[®]

Super A (7-hydroxybutynyl-2-amino-8-aza-7-deazaadenosine) is a duplexstabilizing nucleoside analogue. The Super A:T base pair is much more stable than the natural A:T; it even surpasses the 2-aminoadenosine:T base pair and approaches the stability of the G:C pair. Depending on length and sequence, each Super A incorporation will increase a probe or primer's melting temperature (T_m) by 2.5-5.5°C. Generally, when Super A is located next to a G or C the stabilizing effect is larger (Ref. [14\)](#page-210-0).

Super T®

Super T (5-hydroxybutynyl-2'-deoxyuridine) is another duplex-stabilizing nucleoside analogue which forms a stabilized Super T:A base pair with an average T_m increase of about 1.5-3 \degree C per incorporation, presumably due to an increased stacking interaction between nucleotide bases (Ref. [12,](#page-210-1) [16\)](#page-210-2). The stabilizing effect is highest when several Super Ts are incorporated consecutively.

Figure 6 Structures of Super G, Super A, and Super T with their corresponding complementary base pairs.

Guidelines for incorporating Super A and Super T bases into PCR primers

An example of the typical benefits of primers containing modified bases is shown in [Figure 7](#page-118-0) where they are used to amplify the highly variable *gag* gene region of HIV. Only short sequences are conserved in that region and therefore available for primer design. The unmodified primer pair did not produce any visible PCR products on the agarose gel, whereas the primer pair stabilized by multiple incorporations of Super A and Super T (Forward ACCAAGGAAGC, Reverse CCTTCTGATAATGCTG) produced a good band of product with the expected 240 bp size. Other examples of successful Super A and Super T incorporation can be found in the research literature (Ref. [12,](#page-210-1) [13,](#page-210-3) [14,](#page-210-0) [15,](#page-210-4) [16,](#page-210-2) [17\)](#page-210-5).

Figure 7 Stabilizing moieties of Super A and Super T in short primers improve amplification of highly variable templates.

Cloned HIV variants (gag gene region) were amplified with 0.5 µM each of modified or unmodified primers using JumpStart™ Taq ReadyMix™ (Sigma, St. Louis, MO, USA) using the following cycling parameters: 2 min at 95°C, followed by 50 cycles of 15 s at 95°C and 1 min at 60°C. PCR products were run on a 4% agarose gel. Lanes 2-5 indicate the modified primer pair amplification and lanes 7-10 indicated the unmodified primer pair amplification. Lanes 2 &7 are no template controls. Lanes 3 & 8 contain the pN43 (wild type) plasmid template. Lanes 4 & 9 contain the pN43 – K103N (mutant) plasmid template. Lanes 5 & 10 contain the pN43 – V108I (mutant) plasmid template. Lanes 1 & 6 contain a 100 bp DNA ladder.

It should be noted that PCR efficiency is also affected by factors beyond the thermodynamic properties of primers. DNA polymerase must read through the modified bases without stuttering or pausing, and modified primers have to be efficiently extended. We have conducted experiments and summarized general guidelines for using Super A and Super T in PCR primers:

- Usual PCR T_m requirements should be used to ensure primer annealing.
- Neither Super A nor Super T should be used at the 3' end.
- Both Super A and Super T can be used as the second base from the 3' end.
- Super A should be used sparingly.
- Multiple Super A's should be separated by at least three natural bases or Super T.
- Super T can be used liberally.

Super I^{\circledR}

Super I™ is a nucleoside analogue that contains a quasi-universal nucleobase (7-aminobutynyl-8-aza-7-deazahypoxanthine) with improved hybridization and PCR enhancing properties (Ref. [18\)](#page-210-6) as compared to commonly used 2'-deoxyinosine. It demonstrates improved ambiguity for pairing with A, T and C bases and its base pairing properties can be summarized as follows: X:C~X:A~X:T>X:G. The base pairing strength of Super I with A, T and C is approximately equal or slightly greater (depending on nearest neighbors) than the natural A:T but lower than the G:C, with the Super I:C base pair being slightly more stable than Super I:A and Super I:T. The improvement in PCR performance directly correlated with primers' T_m . Primers with multiple Super I incorporations can be used without significant inhibition of Taq polymerase activity provided the modifications are not used in stretches and are positioned more than 2 bases away from the 3' end.

Figure 8 Possible hydrogen bonding patterns of Super I with natural nucleobases.

Comparison of Super I and 2'-deoxyinosine

We have determined that the stabilities of Super I:A, T and C base pairs are approximately equal or slightly greater (depending on nearest neighbors) than the natural A:T but lower than G:C, with Super I:C being slightly more stable than Super I:A and Super I:T. The beneficial stabilization effect of Super I pairs with A, T and C is especially pronounced for the multiplysubstituted duplexes, which are generally more stable than the respective natural matched duplexes. In contrast, 2'-deoxyinosine forms base pairs with all four natural bases that are weaker than the A:T base pair. Moreover, multiple incorporations of 2'-deoxyinosine render modified duplexes much less stable than the respective natural matched duplexes. The much weaker Super I:G base pair is approximately equivalent to dI:G, both of which significantly destabilize the DNA duplexes. For practical applications such as PCR primer design where maintaining primer melting temperature is imperative, these results imply that single and especially multiple substitutions of A, T and G bases with Super I would be thermodynamically favorable whereas substitutions of cytosines should be avoided.

Super A®

Super A (7-hydroxybutynyl-2-amino-8-aza-7 deazaadenosine) is a duplex-stabilizing nucleoside analogue. The Super A:T base pair is more stable than natural A:T and even surpasses the 2 aminoadenine:T base pair and approaches the stability of the G:C pair. Depending on length and sequence each Super A incorporation will increase the probe or primer's melting temperature (T_m) by 2.5-5.5°C.

Extinction coefficient (260 nm) 8,700 M⁻¹cm⁻¹

Super A 3'-Phosphoramidite

Product number: M830249; CAS #: 2378004-08-9

Super A 5'-Phosphoramidite

Product number: M830250 CAS #: 2378004-04-5

Super G®

но \sqcap ро OH N NH N N O $NH₂$

Super G (8-aza-7-deazaguanosine) eliminates undesired secondary structures inherent to natural G-rich sequences and provides improved hybridization efficiency. It is recommended for use in NA probes and primers to disrupt G stretches.

An additional benefit of Super G for use in fluorogenic hybridization probes is its low fluorescence quenching tendency, which is an intrinsic and often undesired property of natural G. The resultant Super G-modified probes demonstrate higher fluorescence efficiency and, therefore, assay sensitivity.

Extinction coefficient (260 nm) $12,400$ M⁻¹cm⁻¹

Super G 3'-Phosphoramidite

Product number: M830006; CAS #: 500891-26-9

Super G 5'-Phosphoramidite

Product number: M830031; CAS #: 2378004-11-4

Super T®

Super T (5-hydroxybutynyl-2'-deoxyuridine) is another duplex-stabilizing nucleoside analogue which forms a stabilized Super T:A base pair with an average T_m increase of about 2°C per incorporation. The Super T nucleobase can be used liberally in both probe and primer designs with little or no adverse effect on amplification efficiency.

Super T 3'-Phosphoramidite

Product number: M830040; CAS #: 2378004-13-6

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Super T 5'-Phosphoramidite

Product number: M830041; CAS #: 2378004-01-2

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Super I®

Super I is a nucleoside analogue that contains a quasiuniversal nucleobase (7-aminobutynyl-8-aza-7 deazahypoxanthine) with improved hybridization and polymerase chain reaction (PCR) enhancing properties as compared to commonly used 2'-deoxyinosine.

It demonstrates improved ambiguity for pairing with A, T and C bases and its base pairing properties can be summarized as follows: X:C~X:A~X:T>X:G. The base pairing strength of Super I with A, T and C is approximately equal or slightly greater (depending on nearest neighbors) than natural A:T but lower than G:C, with the Super I:C base pair being slightly more stable than Super I:A and Super I:T. The improvement in PCR performance directly correlated with primers' T_m .

Primers with multiple Super I incorporations can be used without noticeable inhibition of Taq polymerase activity provided the modifications are not used in stretches and are positioned more than 2 bases away from the 3' end.

Super I 3'-Phosphoramidite

Product number: M830193; CAS #: 1401110-57-3

Super I 5'-Phosphoramidite

Product number: M830712; CAS #: 2378004-03-4

6. Duplex Intercalators Pyrene Internal Phosphoramidite

Product number: M830711; CAS #: 2378724-62-8

Pyrene-functionalized oligonucleotides are useful tools in fundamental research, diagnostics, and materials science. Their popularity is linked to the ability of pyrenes to function as polarity-sensitive and quenchable fluorophores, excimer-generating units, aromatic stacking moieties, and nucleic acid duplex intercalators (Ref. [28\)](#page-211-0). Pyrene demonstrates a long excited state lifetime, a large Stokes shift, and chemical stability.

Pyrene internal phosphoramidite (M830711) is prepared by coupling of 1-pyrenecarboxylic acid to D-threoninol, a trifunctional linker with an amino group and two hydroxyl groups in a specific steric bond configuration. The pyrene moiety in this combination is a universal base capable of binding equally well with all four natural nucleobases (unpublished data). To achieve optimal backbone configuration the phosphoramidite should be used in conjunction with 5'-nucleoside phosphoramidites for probe synthesis.

The phosphoramidite is suitable for internal as well as terminal incorporation and compatible with standard DNA synthesis and both standard and mild deprotection conditions.

Pyrene-Super I 3'-Phosphoramidite

Product number: M830782; CAS #: 2378004-05-6

7. 3'-Deoxyribopyranosyl(4′→2′) Nucleic Acid

Acronym: p-DNA

3′‐Deoxy‐β-D-ribopyranosyl nucleic acids (p-DNA) are polymers that preferentially pair with p-DNA versus natural DNA sequences (Ref. [29\)](#page-211-1). p-DNAs form antiparallel, exclusively Watson-Crick-paired duplexes that are much stronger than corresponding DNA duplexes.

For some applications, it is essential to be able to design large numbers of nucleic acid sequences with the same T_m that hybridize efficiently and specifically with respective targets while not cross-hybridizing with each other (Ref. [30\)](#page-211-2). These so-called T_m-leveled orthogonal nucleic acids can be designed using p-DNA monomers (Ref. [31\)](#page-211-3). Orthogonal p-DNAs with high duplex stability, low affinity for DNA, and which are not recognized by DNA processing enzymes are useful for labeling, barcoding, or anchoring multiple DNA-containing substrates co-existing in one mixture, on one array, or on a lateral flow strip.

p-DNA monomers are available as 4'-DMT, 2'-phosphoramidites and are fully compatible with standard DNA synthesis. In particular, synthesis of chimeric DNA – p-DNA can be achieved via standard synthesis.

p-DNA T Phosphoramidite

Product number: M830779; CAS #: 256648-34-7

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p-DNA A Phosphoramidite

Product number: M830776; CAS #: 256648-33-6

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p-DNA C Phosphoramidite

Product number: M830777; CAS #: 2378004-09-0

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p-DNA G Phosphoramidite

Product number: M830778; CAS #: 457880-65-8

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8. Non-constrained Nucleic Acid

Acronym: NNA

Non-constrained Nucleic Acid (NNA) is an artificial acyclic phosphodiester DNA analogue composed of stereoisomerically pure (S,S)-2-(nucleobasemethyl)butane-1,3-diols.

The (S,S)-stereoisomer of NNA forms homoduplexes of greater stability than DNA oligonucleotides of the same sequences and does not hybridize with natural DNA to form heteroduplexes (Ref. [32\)](#page-211-4). For a 10-12-mer, on average, the NNA homoduplexes' melting temperature (T_m) is 8°C higher than the respective DNA [\(Figure 9\)](#page-140-0) and about 8°C lower than the p-DNA duplex depending on nucleobase composition.

Figure 9 Melting temperature (T_m) comparison of NNA, DNA and p-DNA duplexes of different nucleobase composition.

NNA, as a tuned down alternative to p-DNA, is also not recognizable by DNA processing enzymes and is useful for labeling, barcoding or anchoring of multiple DNA-containing substrates co-existing in one mixture, array, or lateral flow strip.

NNA T Phosphoramidite

Product number: M830775; CAS #: 2378004-25-0

NNA A Phosphoramidite

Product number: M830772; CAS #: 2378004-35-2

NNA C Phosphoramidite

Product number: M830773; CAS #: 2378004-44-3

NNA G Phosphoramidite

Product number: M830774; CAS #: 2378004-41-0

9. CDPI3 MGB™-Oligonucleotide conjugates and their applications

The tripeptide of dihydropyrroloindole-carboxylate (CDPI3) (Ref. [19\)](#page-211-0) [\(Figure](#page-145-0) [10\)](#page-145-0) is a minor groove binding (MGB) moiety derived from the natural product CC-1065 with strong DNA binding properties. CDPI3 MGB is a crescent-shaped molecule which binds isohelically within the B-form DNA minor groove.

Figure 10 1,2-Dihydro-(*3H*)-pyrrolo[3,2-e]indole-7-carboxylate tripeptide (CDPI3).

The reversible binding is mediated via hydrophobic and van der Waals interactions between the MGB and the floor of the groove. The MGB moiety occupies a region of duplex DNA approximately 5 bases long [\(Figure](#page-145-1) [11\)](#page-145-1) and binds to both A/T and G/C rich sequences with association constants of K_a ~1x10⁷ M⁻¹ and K_a ~1x10⁵ M⁻¹, respectively.

Figure 11 Connolly surface representation of a decamer DNA duplex formed between an MGB-ODN and its complement (Ref[. 20\)](#page-211-1). Red represents the MGB and linker moieties, blue is the MGB-labeled strand and light gray is the complement strand.

Synthetic oligonucleotides (ODNs) with covalently attached CDPI3 MGB moieties were first introduced in 1995 by Lukhtanov et al. (Ref. [21\)](#page-211-2). It has since been shown that such ODNs have enhanced DNA affinity and have improved the hybridization properties of sequence-specific DNA probes. Short MGB-oligonucleotides hybridize with single-stranded DNA to give more stable DNA duplexes than unmodified ODNs of similar length. For example, a DNA duplex formed between the CDPI $_3$ MGB-(dT) $_8$ conjugate and poly(dA) template has a melting temperature (T_m) that is 44°C higher than an unmodified duplex. Mismatch discrimination of short MGB-ODNs is also enhanced versus unmodified oligonucleotides. It was demonstrated that mismatches under the MGB binding region were more easily discriminated for a 15-mer MGB-ODN versus an unmodified ODN with up to a 3-fold increase in free energy difference (Ref. [22\)](#page-211-3).

The tethered CDPI3 MGB moiety has a preference for an A/T rich, B-form DNA duplex but is capable of binding to DNA duplexes with mixed sequences as well as to some modified backbone nucleic acids, as investigated by Kutyavin et al. (Ref[. 23\)](#page-211-4). The preference for A/T sequences has a useful practical application—it is well known that DNA duplex T_m depends on A/T and G/C content with A/T rich sequences being much less stable. CDPI₃ MGB tethering significantly reduces the difference, such that probes of equal length have similar T_m values regardless of base composition.

Applications

1. Arrest of primer extension and PCR blockers

CDPI3 MGB-ODN conjugates were investigated for potential use as antigene agents via the inhibition of DNA polymerase (Ref[. 24\)](#page-211-5). The study, which was done in the context of a single-stranded DNA phage, demonstrated that T7 DNA polymerase was physically blocked when a complementary 16-mer 5'- CDPI3 MGB-ODN was hybridized to a downstream site. Blockage was abolished when a single mismatch was introduced. A 16-mer with 3'-CDPI3 MGB moiety also failed to arrest primer extension. The exceptional efficiency of the primer extension arrest was attributed to DNA polymerase's inability to displace the 5' end of the duplex super-stabilized with the CDPI₃ MGB group.

It has since been shown that 5'-CDPI3 MGB-ODNs are able to arrest Taq DNA polymerase and therefore can be used at PCR temperatures as well. This came as a surprise since it was expected that the 5' exonuclease Taq polymerase activity would degrade such duplexes. Evidently, 5'-MGB labeling makes ODNs resistant to 5' exonuclease digestion. Such ODNs can

be used as PCR blockers to prevent amplification of selected DNA sequences.

2. Short and fluorogenic PCR primers

Efficient priming of PCR was demonstrated with 5'-MGB-ODNs as short as 8-mers using modified (touch-down) PCR cycling conditions or 10-16-mers using a regular PCR cycle (Ref. [25\)](#page-211-6). The PCR was shown to produce specific amplification products of expected size. The reduced-length primers were suggested for use for PCR amplification of viral sequences which possess a high degree of variability or techniques such as gene hunting and differential display which amplify multiple sequences using short primer pairs.

5'-MGB-primers which also have an attached 5'-fluorophore are able to quench the dye fluorescence via the photo-induced electron transfer (PET) mechanism, depicted i[n Figure](#page-147-0) 12. Such primers are significantly quenched in a single strand state but become highly fluorescent when incorporated into the PCR amplicon allowing for detection of target amplification.

3. Real-time PCR probes

The stronger binding of MGB-ODNs versus unmodified ODNs allows for more stringent hybridization conditions to be used in DNA hybridization assays. Short MGB-ODNs with improved mismatch discrimination are especially useful in PCR assays since they bind efficiently and specifically during the high-temperature primer extension cycle. Several types of realtime PCR probes that utilize the MGB moiety have been developed.

• MGB TaqMan® probes (Ref. [22\)](#page-211-3) have CDPI3 MGB-Quencher and Fluorophore tethered to the 3' and 5' ends, respectively. Provided that specific sequence is present in the target DNA, the TaqMan probes are degraded by Taq polymerase during PCR releasing unquenched fluorophore.

- MGB Eclipse® hybridization probes (Ref. [13\)](#page-172-0) have the CDPI3 MGB-Quencher and Fluorophore attached at the 5' and 3' ends, respectively. They are non-degradable and their fluorescence is strongly increased upon the probes' hybridization to amplified targets during the annealing step.
- MGB Pleiades® probes (Ref. [8\)](#page-210-0) are also non-degradable hybridization probes. They have the CDPI₃ MGB-Fluorophore and Quencher moieties tethered to the 5' and 3' ends, respectively. These probes utilize the unique dual fluorescence quenching mechanism to significantly reduce background fluorescence and improve signal-to-background ratio.
- MGB FRET probes are similar to MGB Pleiades but contain an additional (donor) fluorophore conjugated to an internal nucleobase. MGB FRET probes allow for multicolor detection with a single wavelength excitation. MGB FRET probes offer high signalto-background ratios due to low background fluorescence via the dual quenching mechanism.

Figure 13 Putative mechanisms of action of MGB TaqMan (A), MGB Eclipse (B), MGB Pleiades (C) and MGB FRET (D) probes.

4. miRNA Inhibitors

In a recent US patent (Ref. [26\)](#page-211-7), it was disclosed that MGB-ODNs with a 2'- OMe sugar-phosphate nucleic acid backbone are highly efficient and specific miRNA inhibitors. The inhibition is more pronounced when the minor groove binder is tethered to the 5'end of a miRNA inhibitor. The mechanism of this effect is not fully understood but could be attributed to the ability of 5'-MGB moiety to stabilize nucleic acid duplex and block enzymatic activities.

MGB-ODN-based miRNA inhibitors have also shown improved cellular uptake and promise as pharmaceuticals by modulating gene expression (Ref. [27\)](#page-211-8).

CDPI3 Minor Groove Binder

Acronym: CDPI3 MGB

CDPI3 MGB is an efficient duplexstabilizing agent. This ligand improves background fluorescence and mismatch discrimination abilities of DNA probes. When attached to the 5'-end of a probe it inhibits the 5' \rightarrow 3' exonuclease activity of DNA polymerases.

**Measured for T10-CDPI3 conjugate (50 mM Tris-HCl pH 8.5, 20°C) **CDPI3 contribution only*

****Suitable for ABI 3900 DNA synthesizer*

CDPI3 PFP ester

Product number: M830187

**Purity of this active derivative is verified by HPLC & NMR and usually exceeds 90%. Some loss of activity, however, may occur during packaging and storage, especially for small (1- 10 mg) unit sizes.*

CDPI3 MGB Terminal Phosphoramidite

Product number: M830252

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CDPI3 MGB CPG

Product number: M830739

CDPI3 MGB Polystyrene

Product numbers: M830203 (bulk), M100103 (columns)

CDPI2 PFP ester

Product numbers: M830790

**Purity of this active derivative is verified by HPLC & NMR and usually exceeds 90%. Some loss of activity, however, may occur during packaging and storage, especially for small (1- 10 mg) unit sizes.*

CDPI4 (SO3H)² PFP ester

Product numbers: M830188

**Purity of this active derivative is verified by HPLC & NMR and usually exceeds 90%. Some loss of activity, however, may occur during packaging and storage, especially for small (1- 10 mg) unit sizes.*

10. PBI3 MGB™-Oligonucleotide conjugates

Piperazino-tris-benzimidazole (PBI3) [\(Figure 14\)](#page-157-0) is a DNA minor groove binder derived from the anthelmintic compound Hoechst 33258. It occupies a region of 5 to 6 base pairs in a DNA duplex and preferentially binds to A/T rich sequences.

Figure 14 Structure of PBI₃ minor groove binder

Similar to CDPI₃, oligonucleotides with a conjugated PBI₃ moiety demonstrate enhanced hybridization properties (table below). While both types of MGB prefer A/T rich sequences, the degree of stabilization of such duplexes by PBI₃ is substantially higher, on average by 10° C for a 12-mer sequence with an A/T rich 5-6 base MGB binding region.

Comparison of DNA duplex stabilization (duplex T_m $(°C)$ **) by conjugated CDPI3 and PBI3 minor groove binders**

**Also contained FAM-HEG (M830100) between MGB and 5'-end. Highlighted are probable MGB binding regions.*

Stabilization of G/C rich sequences is comparably modest for both MGB types, 1-5°C for a 12-mer sequence.

Another aspect of an MGB conjugate is the ability to distinguish between fully (matched) and partially (mismatched) complementary duplexes. With regards to match versus mismatch, both CDPI3 and PBI3 demonstrate similar performance. On average, about 11°C duplex destabilization (T_m decrease) is observed in a 12-mer sequence when a single mismatched base pair is introduced.

Comparison of mismatch discrimination by conjugated CDPI3 and PBI3 minor groove binders

Despite the similarities, PBI3 MGB is an attractive alternative to CDPI3 when duplex hyper-stabilization is required, for instance, for a sequence specific polymerase arrest (PCR blocking).

Another distinctive feature of the PBI₃ moiety is its fluorescence properties. Its fluorescence efficiency increases significantly upon binding with a double-stranded DNA. In the case of a PBI3-oligonucleotide conjugate, fluorescence increase is triggered by the conjugate hybridization with a complementary target. In principle, this allows for a very simple hybridization probe design that requires no additional fluorophore or a quencher.

Optical properties of conjugated PBI3

Measured for T8-PBI3 conjugate (50 mM Tris-HCl pH 8.5, 20°C) **Measured for 200 nM T8-PBI3 conjugate (10 mM Tris-HCl pH 8.9, 40 mM NaCl, 5 mM MgCl2) at 20°C in the presence of 400 nM A8C complement *PBI3 contribution only*

*****Suitable for ABI 3900 DNA synthesizer*

PBI3 MGB PFP ester

Product number: M830795

**Purity of this active derivative is verified by HPLC & NMR and usually exceeds 90%. Some loss of activity, however, may occur during packaging and storage, especially for small (1- 10 mg) unit sizes.*

Available documents CoA, conjugation protocols

PBI3 MGB CPG

Product number: M830796

PBI3 MGB Polystyrene

Product numbers: M830797 (bulk), M100243 (columns)

11. Eclipse Dark Quencher-MGB synthesis supports

Acronym: EDQ-MGB

EDQ-MGB CPG and polystyrene supports offer the most straightforward way of preparing MGB TaqMan and MGB Eclipse probes [\(Figure 13A](#page-148-0) and [Figure 13B](#page-148-0)). When used in combination with 3'-nucleoside phosphoramidites and a fluorophore phosphoramidite at the last step of the synthesis, the supports will yield a 5'-fluorophore, 3'-EDQ-MGB probe, which is the essence of the MGB TaqMan probe's structure. Alternatively, the use of 5'-nucleoside phosphoramidites will afford a 3'-fluorophore, 5'- EDQ-MGB probe known as MGB Eclipse. MGB TaqMan probes generate fluorescence signal during PCR amplification via 5'-exonuclease degradation in the presence of complementary targets. MGB Eclipse probes, on the other hand, are 5'-exonuclease resistant and remain annealed to complementary targets to generate fluorescent signal. Both types of probes benefit from the presence of the duplex-stabilizing MGB moiety which allows for shorter probes with improved mismatch discrimination and reduced background fluorescence.

**Measured for T8-EDQ-MGB conjugate (50 mM Tris-HCl pH 8.5, 20°C) **EDQ-MGB contribution only*

EDQ-MGB CPG

Product number: M830394 (500 Å), M830394-1000 (1000 Å)

EDQ-MGB Polystyrene

Product number: M830385 (bulk), M130008 (columns)

DSQ is a single molecule that acts as an MGB and a universal fluorescence quencher at the same time. It was designed by converting a part of the original CDPI3 MGB molecule into a quencher without compromising the MGB duplex-stabilizing properties. DSQ was developed to improve MGB TaqMan probe chemistry. It has a number of advantages over the EDQ-MGB chemistry. It eliminates the need for a separate MGB and quencher and streamlines probe manufacturing. More importantly, DSQ improves DNA duplex stabilization [\(Figure 16\)](#page-168-0), fluorescence quenching [\(Figure 15](#page-167-0) and [Figure 17\)](#page-168-1) and signal-to-background ratio [\(Figure 17\)](#page-168-1).

Figure 15 Comparison of background fluorescence for unhybridized FAM-labeled EDQ-MGB and DSQ probes. Examples include probes specific for BK virus (BK), cytomegalovirus (CMV), and Epstein-Barr Virus (EBV).

Figure 16 Comparison of melting temperatures of DSQ and EDQ-MGB probes.

On average, DSQ-probe/target duplexes are 2°C (range -2 to +5°C) more stable than the respective MGB-probe/target duplexes [\(Figure 16\)](#page-168-0).

When compared to MGB TaqMan, DSQ TaqMan probes show a 20-50% reduction of baseline fluorescence, comparable amplification signals (and Ct values) and 10-100% increase of signal-to-baseline ratio [\(Figure 17\)](#page-168-1).

Figure 17 Comparison of real-time PCR baseline fluorescence and signal-tobaseline ratios of DSQ- and EDQ-MGB-modified probes.

Optical properties of conjugated DSQ

**Measured for T8-DSQ conjugate (50 mM Tris-HCl pH 8.5, 20°C) **DSQ contribution only*

DSQ CPG

Product number: M830762

DSQ Polystyrene

Product number: M830756 (bulk), M100501 (columns)

Capacity by DMT loading $\geq 10 \mu \text{mol/g}$ Purity of test oligo (reverse phase HPLC) \geq 65%
Recommended coupling conditions N/A Recommended coupling conditions Recommended deprotection conditions 30% NH₄OH/EtOH (3:1; v:v), 16 h at 55°C or 2 h at 70°C Recommended storage conditions/Stability 2-8°C, dry/ 5 years Available documents Certificate of analysis

13. Fluorophore-MGB (CDPI3) Polystyrene supports

Fluorophore-MGB polystyrene supports have been developed for simultaneous incorporation of a fluorophore and the MGB residue either at the 3' or 5' end of an oligonucleotide. When used in combination with 5'-nucleoside phosphoramidites and a non-fluorescent quencher, these supports yield 5'-fluorophore-MGB, 3'-quencher oligonucleotide probes known as MGB Pleiades probes [\(Figure 13C](#page-148-0)). MGB Pleiades are hybridization-triggered fluorogenic probes, which are resistant to 5' exonuclease degradation and thus suitable for post-PCR melting curve analysis. Owing to the efficiency of a dual quenching mechanism, mediated by both the MGB and a non-fluorescent quencher, Pleiades probes possess a very low background fluorescence in the absence of a target. Fluorescence is restored upon probe hybridization with complementary target with excellent signal-to-background ratios. Additionally, fluorophores are separated from the MGB group by the hexa-ethylene glycol (HEG) spacer for improved hybridization-dependent fluorescence.

EB380-MGB Polystyrene

Product number: M830740 (bulk), M100431 (columns)

FAM-MGB Polystyrene

Product number: M830693 (bulk), M100185 (columns)

AP525-MGB Polystyrene Product number: M830692 (bulk), M100186 (columns) HN N O HN N o″ N H N NH \gtrsim O O O $\mathsf{o}_{\!\scriptscriptstyle \times\! \scriptscriptstyle \times} \mathsf{o}$ O be O $P - O$ O O N ODMT O \sim o \sim o \sim o \sim o O N HN \circledast or \circledast or O O o _{Cl} O P O o
‼o NH O Cl HN O CF_3 $CF₃$ O

AP559-MGB Polystyrene

Product number: M830689 (bulk), M100181 (columns)

AP593-MGB Polystyrene

Product number: M830685 (bulk), M100135 (columns)

AP639-MGB Polystyrene

Product number: M830738 (bulk), M100293 (columns)

AP642-MGB Polystyrene Product number: M830200 (bulk), M100153 (columns)

AP662-MGB Polystyrene Product number: M830695 (bulk), M100250 (columns)

AP690-MGB Polystyrene

Product number: M830754 (bulk), M100499 (columns)

Cholesterol Phosphoramidite

Product number: M830785; CAS #: 143723-64-2

15. Amine-modifiers, Linkers, and Spacers

Benzaldehyde Phosphoramidite

Product number: M830547; CAS #: 433684-36-7

Benzaldehyde phosphoramidite is suitable for the introduction of an aromatic aldehyde at the 5' or 3' end of an oligonucleotide. The cyclic acetal provides a hydrophobic handle for reverse phase HPLC purification and can be removed after HPLC by treatment with 80% acetic acid. Alternatively, the acetal protection is removed during the standard on-line detritylation step.

Aromatic aldehydes are mild electrophiles, which are capable of forming Schiff base type adducts with various nucleophiles, e.g., amines, hydrazines or hydrazides (Ref[. 33\)](#page-211-0).

DMT-Hexanol Diglycolate CPG

Product number: M830758

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DMT-Hexanol Diglycolate Polystyrene

Product number: M126137 (bulk), M100457 (columns)

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TFA-Aminopropyl-dU 3'-Phosphoramidite

Product number: M830780; CAS #: 134140-86-6

TFA-Aminopropyl-dU 5'-Phosphoramidite

Product number: M830715; CAS #: 1146714-75-1

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TFA-Aminopropyl-Super G 3'-Phosphoramidite

Product number: M830784; CAS #: 872840-74-9

Fmoc-Hydroxyprolinol Phosphoramidite

Product number: M830769; CAS #: 2378004-37-4

Fmoc-Hydroxyprolinol CPG

Product number: M830768

Fmoc-Hydroxyprolinol Polystyrene

Product number: M830770 (bulk), M100510 (columns)

Fmoc-Aminohexanediol CPG

Product number: M830765

Fmoc-Aminohexanediol Polystyrene

Product number: M830766 (bulk), M100511 (columns)

Fmoc-HEG Phosphoramidite

Product number: M830767; CAS #: 2378004-40-9

Fmoc-HEG-MGB Polystyrene

Product number: M830798 (bulk), M100508 (columns)

Appendix A. Determination of the relative hydrophobicities of conjugated dyes using C18 column chromatography

Figure 18 HPLC analysis of a mixture of fluorescent dye calibrators.

1 - T8-RR (M830731); 2 - T8-AP525 (M300587); 3 - T8-FAM-HEG (M300586); 4 - T8-EB380-HEG (M300585); 5 - T8-YY (M830730); 6 - T8-AP559 (M301430); 7 - T8-AP570 (M830700) ; 8 - T8- AP593 (M300392) ; 9 - T8-AP642 (M300589) ; 10 - T8-AP662 (M830724) ; 11 - T8-AP680 (M300753).

Column and HPLC conditions: C18 Luna (4.6x100 mm), Phenomenex (00D-4041-E0). Sample loop – 20 µL. Flow rate – 1ml/min, Solvent A – 0.1 M triethylammonium acetate pH 7.5. Solvent B – CH3CN. Gradient – 0-40% B in 19 min. Detection – 260 nm.

Appendix B. Determination of molar extinction coefficients of conjugated fluorophores and quenchers

The determination of extinction coefficient for a conjugated dye may be challenging due to the sensitivity of absorption and fluorescence spectra to the dye's environment and potentially complex dye-conjugate interactions. In the case of oligonucleotide conjugates, for instance, dyes may interact with nucleobases as well as other conjugated moieties. To take into account these effects, an optimal measurement procedure should employ a dye-oligonucleotide conjugate. However, determination of the dyeoligonucleotide concentration by pure conjugate mass, which is required for the direct extinction coefficient measurement, is often impractical. We have overcome this problem by using a concentration-independent approach which utilizes the known extinction coefficients of oligothymidylates and the dye absorption as an internal reference. In this approach, a series of dye-oligothymidylate conjugates is prepared with a variable number of thymidine units. The molar extinction coefficient for oligothymidylate can be described by a simple formula E260 = 8100*n* + 600, where *n* is the number of thymidine residues. Assuming that the dye conjugation does not affect this dependence, the combined extinction coefficient for dye-oligoT conjugate can be represented as E260 (conjugate) $= 8100n + 600 + E_{260}(dye)$. To obtain E₂₆₀(dye), one must measure A₂₆₀ (260) nm absorption), Aλmax(dye) (absorption at the dye maximum) and plot A_{260} (conjugate)/ $A_{\lambda max}$ (dye) values over the number of thymidine units. These data are then analyzed by linear regression to generate a line of best fit and its corresponding equation $y = ax + b$. Since T oligomers' slope parameter *a* equals 8100, the equation can be rewritten as *y* = 8100*n* + *b*/*a*8100*.*

The procedure is illustrated in [Figure](#page-200-0) 19. In this example, the described approach was used to determine an extinction coefficient for a T_nconjugated AP593 dye.

It should be noted that the success of this method relies on several assumptions and requirements listed below:

- 1) Dye-oligonucleotide interactions are independent on the length of the oligomers;
- 2) All conjugates must be carefully purified to ensure accurate A₂₆₀ measurements;
- 3) The linear fit of all data points should have a high (>0.99) R^2 value to ensure accurate y-intercept determination.

C E₂₆₀(dye) = 0.1746 * 8100 / 0.0698 – 600 = 19661.6 (M⁻¹cm⁻¹)

Figure 19 Extinction coefficient (E₂₆₀) determination for the AP593 dye conjugated to oligothymidylates.

(A) Normalized (at 593 nm) absorption spectra of T12-, T10-, T8- and T6-AP593 conjugates. (B) Line of best fit (with equation and R2 value) for the normalized A260 values versus number of thymidine units. (C) Calculation of the E₂₆₀(dye) coefficient using the parameters a and b from the best fit linear equation. The y-intercept (b/a8100) of this line represents $E_{260}(dye) + 600$ *. Therefore, E260(dye) =* b*8100/*a *– 600.*

Appendix C. Effect of pH on dye fluorescence

Experimental conditions: Instrument: Varian Cary Eclipse; 5 nm slit width; Buffer: 40 mM sodium phosphate; Temperature: 20°*C.*

Appendix D. Photostability of fluorescent dyes

Figure 21 Relative photostability of fluorescent dyes.

0 0.5 1 1.5 2 2.5 3 3.5 4 **lamp exposure time, hours**

 $-AP690$

0

500 nM dye calibrator solutions in TE (pH 8.0) were exposed to a 250 W incandescent bulb at a distance of 20 cm, using a water bath to maintain temperature at 20-35°C.

Note: AP559 and AP570 data partly obscured by AP593; AP639 data partly obscured by AP690.

Appendix E. Fluorescence temperature dependence

Figure 22 Effect of temperature increase (from 20°C to 80°C) on dye fluorescence. *Concentration: 100 nM dye calibrator in 50 mM Tris-HCl (pH 8.0)*

QUICK REFERENCE GUIDE: FLUORESCENT DYES

*See Appendix A

**See Appendix D

QUICK GUIDE FOR OLIGONUCLEOTIDE SYNTHESIS: PHOSPHORAMIDITES

**Optimized for ABI 3900 DNA Synthesizer. Other instruments' coupling conditions may vary.*

QUICK GUIDE FOR OLIGONUCLEOTIDE SYNTHESIS: SYNTHESIS SUPPORTS

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PRODUCT INDEX

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Patents

The Minor groove binder-, dye- and modified base- reagents and their applications are covered by one or more of U.S. Patents Nos.: US6,972,339, US7,112,684, US7,348,146, US7,381,818, US7,541,454, US7,553,643, US7,582,739; US7,601,851, US7,662,942, US7,671,218, US7,767,834, US7,851,606, US7,897,736, US8,008,522, US8,067,177; US8,163,910, US7,205,105, US8,389,745, US6,949,367, US8,465,921, US7,045,610, US7,368,549, US7,485,442, US6,683,173, US7,751,982, US7,715,989, US6,660,845, US6,972,328, US6,962,991, US7,759,126, US9,085,800, US9,056,887, US9,169,256, US9,328,384; US9,334,495, US9,932,643, US9,677,142 US9,988,670, US10,127,349, US10,266,903, US10,590,474, US 677,728, US10,738,346, US10,890,529, US10,975,423, US11,155,713, US11,242,554, US11,320,376, Foreign Patents Nos.: EP3592757A2, EP2971106B1, EP3230468B1, EP3230468B1, EP2714939B1, EP2801626B1, CA2835283C, JP6196968B2, EP2547769B1, EP2997161B1, EP2689031B1, EP2736916B1, JP6182529B2, EP2689031B1, EP1781675B1, JP5214967B2, JP4510626B2, JP4942484B2, EP1687609B1, EP1789587B1, EP01781675B1, JP05214967B2, EP1430147B1, JP4510626B2, EP1687609B1, JP4942484B2, EP1789587B1 , JP4510626B2, EP3668884B1, EP3592757A2 as well as US and international applications that are currently pending.

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